



A Soluble Polymer-Bound Approach to the Sharpless Catalytic Asymmetric Dihydroxylation (AD) Reaction: Preparation and Application of a [(DHQD)₂PHAL-PEG-OMe] Ligand

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Abstract: The synthesis of a soluble polymer-bound (DHQD)₂PHAL ligand **1** and its successful application in the catalytic asymmetric dihydroxylation reaction are described. The results highlight that the soluble polymer-bound ligand approach offers all the benefits associated with insoluble supports but with the key advantages of increased reactivity and enantioselectivity inherent with solution-phase methodology. © 1997 Elsevier Science Ltd. All rights reserved.

Since its discovery in 1988,¹ Sharpless' catalytic asymmetric dihydroxylation (AD) of olefins has been continuously refined by both a development of better ligands and improvements in the secondary oxidant/solvent system.² In parallel, Sharpless³ and other groups⁴ have investigated immobilization of these expensive ligands onto insoluble polymer supports so as to aid in their recovery. While this strategy provides a simple but elegant way for automating the AD reaction, it has a number of limitations including prolonged reaction times and, more importantly, a reduction in enantioselectivity.

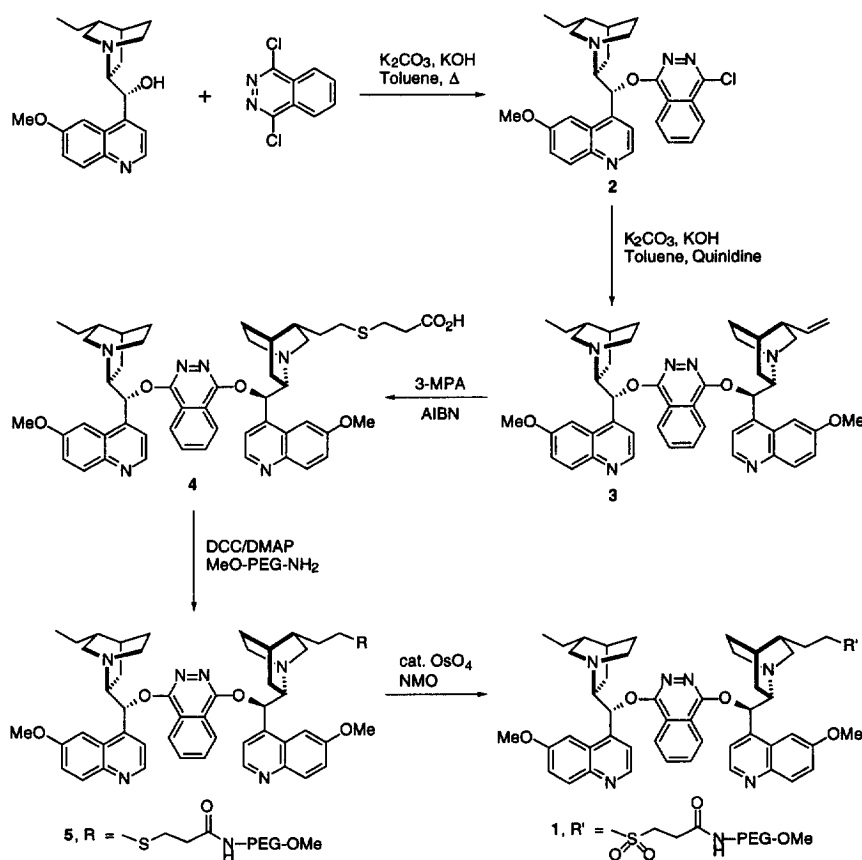
In an effort to circumvent the problems associated with heterogeneous reactions, we recently reported a homogeneous extension to the AD reaction using a soluble polymer, poly(ethylene glycol) mono-methyl ether (MeO-PEG), bound cinchona alkaloid ligand.⁵ This liquid-phase methodology⁶ provided all of the advantages that an insoluble polymer has to offer, while also being as effective as the free ligand in terms of both reactivity and enantioselectivity. However, in our preliminary report, the ligand utilized was not the most effective in terms of enantioselectivity and reactivity for the solution-phase AD reaction. Therefore in an attempt to improve our liquid-phase approach to the AD reaction we now report the synthesis of a (DHQD)₂PHAL ligand bound to MeO-PEG-NH₂ and its successful use in the AD reaction of various olefins.

The five step synthetic route to **1** is outlined in Scheme 1.⁷ In the first step, a mixture of dihydroquinidine, 1,4-dichlorophthalazine, KOH, and K₂CO₃ in dry toluene are refluxed, with a concurrent azeotropic removal of water, to give the mono-substituted chlorophthalazine **2** which upon similar transformation with quinidine provided the di-substituted phthalazine **3**.⁸ The heating of **3** and 3-mercaptopropionic acid (3-MPA) in the presence of 2,2'-azobisisobutyronitrile (AIBN) in benzene (70 °C) allowed the isolation of **4** as a tan precipitate.⁹ The acid **4** was coupled to MeO-PEG-NH₂ in the presence of N,N-dimethylaminopyridine (DMAP) and 1,3-dicyclohexylcarbodiimide (DCC) in methylene chloride (DCM).¹⁰ After the reaction was complete, the insoluble urea side-product was removed by filtration and the pegylated ligand **5** was isolated from the reaction mixture by precipitation following a slow addition of diethyl

ether. The sulfide **5** was oxidized to the desired sulfone **1** by a mixture of OsO_4 /*N*-methylmorpholine-*N*-oxide (NMO) [in acetone/water (v/v, 2/1)].¹¹

Ligand **1** was completely soluble either in *t*-butanol/water or acetone/water solvent systems allowing the study of homogeneous AD reactions. The AD reaction results of **1** with various olefins are shown in Table 1 and a number of features are noteworthy. First, it is evident that the *t*-Butanol/water solvent produces considerably higher *ees* for all olefins tested consistent with previous reports.⁴ Second, in terms of both reaction time and enantioselectivity, ligand **1** accelerated AD reactions are comparable to its free ligand counterpart, strongly suggesting that the MeO-PEG backbone does not adversely alter either asymmetric induction or the rate of formation of the osmium-ligand-olefin ternary complex. Finally, the ligand **1** can be isolated in virtually quantitative yield by precipitation using diethyl ether.⁵


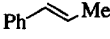
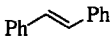
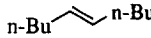
Scheme 1



Combined with our previous report,⁵ the present results demonstrate that MeO-PEG bound ligands behave in a similar fashion to unimmobilized ligands in the AD reaction. What makes this finding even more

impressive is that this soluble polymer approach provides the added convenience of ligand recovery and product isolation. We believe that this methodology for ligand/catalyst recovery should be useful and applicable to other catalytic reactions and will be reported in due course.

Table 1. Catalytic Asymmetric Dihydroxylation Reactions using Ligand 1.^a

Entry	Olefin	Oxidant	Yield (%)	ee (%)
1		NMO	87	72
2		K ₃ FeCN ₆	88	98 (97) ^b
3		NMO	87	91
4		K ₃ FeCN ₆	83	99 (99) ^b
5		NMO	98	94
6		K ₃ FeCN ₆	95	99 (>99) ^b
7		NMO	84	80
8		K ₃ FeCN ₆	80	97 (97) ^b

^aFor NMO system, the molar ratio of olefin/OsO₄/ligand = 1/0.04/0.1, and for K₃FeCN₆ system, the molar ratio was 1/0.005/0.1. For other details, see references 3 and 5. ^bNumber in parenthesis represents results for a free ligand (DHQD)₂PHAL from reference 12.

Acknowledgements This work was supported in part by the R. W. Johnson Pharmaceutical Research Institute and The Skaggs Institute for Chemical Biology at The Scripps Research Institute.

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7. **2**: ¹NMR (300 MHz, CDCl₃) δ 0.91 (t, J = 6.7 Hz, 3H), 1.40-1.70 (m, 6H), 1.77 (m, 1H), 2.12 (m, 1H), 2.65-3.00 (m, 4H), 3.43-3.58 (m, 1H), 3.99 (s, 3H), 7.27 (d, J = 7.0 Hz, 1H), 7.34 (dd, J = 9.3 & 2.8 Hz, 1H), 7.44 (d, J = 4.5 Hz, 1H), 7.62 (d, J = 2.7 Hz, 1H), 7.96 (d, J = 9.2 Hz, 1H), 8.00 (m, 2H), 8.13-8.20 (m, 1H), 8.33-8.42 (m, 1H), 8.64 (d, J = 4.5 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 11.9, 23.5, 25.4, 26.1, 27.1, 37.3, 49.9, 50.9, 55.5, 59.9, 101.7, 118.5, 121.4, 121.8, 122.8, 122.9, 125.3, 127.7, 128.2, 131.6, 133.0, 133.3, 144.0, 144.7, 147.0, 147.2, 150.5, 157.7; HRMS (FAB⁺) calcd for [C₂₈H₂₉ClN₄O₂ + H⁺] = 489.2057, found = 489.2051.
- 3**: ¹NMR (300 MHz, CDCl₃) δ 0.78 (t, J = 6.7 Hz, 3H), 1.20-1.65 (m, 10H), 1.68 (m, 1H), 1.85-2.30 (m, 3H), 2.55-3.00 (m, 8H), 3.39 (m, 2H), 3.88 (s, 6H), 4.97 (m, 2H), 5.90 (m, 1H), 6.95 (d, J = 6.5 Hz, 1H), 7.01 (d, J = 5.9 Hz, 1H), 7.34 (m, 2H), 7.40 (d, J = 4.6 Hz, 1H), 7.42 (d, J = 4.6 Hz, 1H), 7.52 (d, J = 2.7 Hz, 1H), 7.54 (d, J = 2.6 Hz, 1H), 7.91 (m, 2H), 7.96 (d, J = 9.2 Hz, 1H), 7.97 (d, J = 9.2 Hz, 1H), 8.31 (m, 2H), 8.61 (d, J = 4.5 Hz, 1H), 8.62 (d, J = 4.5 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 11.8, 23.2, 23.4, 25.2, 26.0, 26.2, 26.4, 27.2, 27.7, 37.3, 39.6, 49.4, 49.8, 49.9, 50.8, 55.5, 60.0, 60.2, 76.0, 77.2, 101.9, 102.0, 114.6, 118.2, 118.4, 121.7, 121.8, 122.3, 122.7, 123.0, 127.3, 128.1, 129.0, 131.4, 131.5, 132.0, 132.1, 140.3, 144.6, 144.8, 144.9, 147.3, 156.3, 156.4, 157.5, 157.6; HRMS (FAB⁺) calcd for [C₄₈H₅₂N₆O₄ + H⁺] = 777.4128, found 777.4104.
- 4**: ¹NMR (300 MHz, CDCl₃) δ 0.86 (t, J = 6.7 Hz, 3H), 1.25-3.25 (m, 30H), 3.50 (m, 2H), 3.89 (s, 3H), 3.97 (s, 3H), 6.60 (broad s, 1H), 7.30 (m, 3H), 7.44 (t, J = 4.4 Hz, 2H), 7.49 (broad s, 1H), 7.53 (d, J = 2.3, 1H), 7.63 (d, J = 2.4 Hz, 1H), 7.88 (m, 2H), 7.95 (d, J = 9.2 Hz, 1H), 7.96 (d, J = 9.2 Hz, 1H), 8.19 (d, J = 8.1 Hz, 1H), 8.27 (d, J = 7.7 Hz, 1H), 8.59 (d, J = 4.6 Hz, 1H), 8.64 (d, J = 4.5 Hz, 1H); HRMS (FAB⁺) calcd for [C₅₁H₅₈N₆O₆S + H⁺] = 883.4216, found 883.4242.
- 5**: ¹NMR (300 MHz, CDCl₃) δ 0.87 (t, J = 6.7 Hz, 3H), 1.25-2.85 (m, 30H), 3.20-3.75 (PEG peaks), 3.84 (t, J = 6.5 Hz, 2H), 3.88 (s, 6H), 6.59 (broad t, 1H), 6.90 (d, J = 6.6 Hz, 1H), 6.97 (d, J = 5.7 Hz, 1H), 7.30 (dd, J = 9.2 & 2.4 Hz, 1H), 7.32 (dd, J = 9.0 & 2.4 Hz, 1H), 7.39 (t, J = 4.6 Hz, 2H), 7.47 (d, J = 2.5 Hz, 1H), 7.51 (d, J = 2.5 Hz, 1H), 7.90 (m, 2H), 7.93 (d, J = 9.0 Hz, 1H), 7.94 (d, J = 9.2 Hz, 1H), 8.28 (m, 2H), 8.59 (d, J = 4.5 Hz, 2H).
- 1**: ¹NMR (300 MHz, CDCl₃) δ 0.79 (3H), 1.10-3.00 (30H), 3.10-3.80 (PEG peaks), 3.80-4.00 (8H), 6.70 (1H), 6.95 (1H), 7.05 (1H), 7.20-7.45 (7H), 7.50 (1H), 7.80-8.10 (4H), 8.28 (1H), 8.37 (1H), 8.55 (2H). Peak multiplicity was not further characterized due to line-broadening.
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(Received in USA 19 December 1996; revised 15 January 1997; accepted 16 January 1997)